

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
1 November 2001 (01.11.2001)

PCT

(10) International Publication Number
WO 01/80920 A2

(51) International Patent Classification⁷: A61L 27/54,
27/30, 31/08, 31/16, 17/14, 15/46, A61F 13/00

(74) Agent: MCKAY-CAREY, Mary, Jane; McKay-Carey
& Company, Suite 2590 Commerce Place, 10155 - 102nd
Street, Edmonton, Alberta T5J 4G8 (CA).

(21) International Application Number: PCT/CA01/00498

(22) International Filing Date: 17 April 2001 (17.04.2001)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
60/197,959 17 April 2000 (17.04.2000) US

(71) Applicant (for all designated States except US): WES-
TAIM BIOMEDICAL CORP. [CA/CA]; 10102-114
Street, Fort Saskatchewan, Alberta T8L 3W4 (CA).

(72) Inventors; and

(75) Inventors/Applicants (for US only): BURRELL, Robert,
Edward [CA/CA]; 52055, R.R. 221, Sherwood Park, Al-
berta T8E 1C6 (CA). YIN, Hua, Qing [CN/CA]; 24 Davy
Crescent, Sherwood Park, Alberta T8H 1P3 (CA). DJO-
KIC, Stojan [CA/CA]; #301, 15825 Beaumaris Road, Ed-
monton, Alberta T5X 5H1 (CA). LANGFORD, Rita, Jo-
hanna, Mary [CA/CA]; 3244 - 36A Avenue, Edmonton,
Alberta T6T 1G3 (CA).

(81) Designated States (national): AE, AG, AL, AM, AT, AU,
AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU,
CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM,
HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK,
LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX,
MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL,
TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM,
KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian
patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European
patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE,
IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF,
CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

— without international search report and to be republished
upon receipt of that report

For two-letter codes and other abbreviations, refer to the "Guid-
ance Notes on Codes and Abbreviations" appearing at the begin-
ning of each regular issue of the PCT Gazette.

(54) Title: ANTIMICROBIAL BIOABSORBABLE MATERIALS

(57) Abstract: The invention provides bioabsorbable materials with antimicrobial coatings or powders which provide an effective and sustainable antimicrobial effect. Specifically, this invention provides bioabsorbable materials comprising a bioabsorbable substrate associated with one or more antimicrobial metals being in a crystalline form characterized by sufficient atomic disorder, such that the bioabsorbable material in contact with an alcohol or water based electrolyte, releases atoms, ion, molecules, or clusters of at least one antimicrobial metal at a concentration sufficient to provide an antimicrobial effect. The one or more antimicrobial metals do not interfere with the bioabsorption of the bioabsorbable material, and do not leave behind particulates larger than 2 μm , as measured 24 hours after the bioabsorbable material has disappeared. Most preferably, the particulate sizing from the coating or powder is sub-micron, that is less than about 1 μm , as measured 24 hours after the bioabsorbable material has disappeared. Particulates are thus sized to avoid deleterious immune responses or toxic effects. Such antimicrobial metals are in the form of a continuous or discontinuous coating, a powder, or a coating on a bioabsorbable powder. The antimicrobial coating is thin, preferably less than 900 nm or more preferably less than 500 nm, and very fine grained, with a grain size (crystallite size) of preferably less than 100 nm, more preferably less than 40 nm, and most preferably less than 20 nm. The antimicrobial coating is formed of an antimicrobial metal, which is overall crystalline, but which is created with atomic disorder, and preferably also having either or both of a) a high oxygen content, as evidenced by a rest potential greater than about 225 mV, more preferably greater than about 250 mV, in 0.15 M Na_2CO_3 against a SCE (standard calomel electrode), or b) discontinuity in the coating. The antimicrobial metal associated with the bioabsorbable substrate may also be in the form of a powder, having a particle size of less than 100 μm , or preferably less than 40 μm , and with a grain size (crystallite size) of preferably less than 100 μm , more preferably less than 40 nm, and most preferably less than 20 nm. Such powders may be prepared as a coating, preferably of the above thickness, onto powdered biocompatible and bioabsorbable substrates; as a nanocrystalline coating and converted into a powder; or as a powder of the antimicrobial metal which is cold worked to impart atomic disorder. Methods of preparing the above antimicrobial materials are thus also provided.

WO 01/80920 A2

1 **"Antimicrobial Bioabsorbable Materials"**

2

3 **FIELD OF THE INVENTION**

4 The invention relates to bioabsorbable materials, which are rendered antimicrobial due to
5 the presence of antimicrobial metals in the form of coatings or powders; processes for their
6 production; and use of same for controlling infection.

7 **BACKGROUND OF THE INVENTION**

8 The risk of acquiring infections from bioabsorbable materials in medical devices is very high.
9 Many medical applications exist for bioabsorbable materials including:

- 10 1) Wound Closures: including for example sutures, staples, adhesives;
11 2) Tissue Repair: including for example meshes for hernia repair;
12 3) Prosthetic Devices: including for example internal bone fixation, physical barrier for guided
13 bone regeneration;
14 4) Tissue Engineering: including for example blood vessels, skin, bone, cartilage, and liver; and
15 5) Controlled Drug Delivery Systems: including for example microcapsules and ion-exchange
16 resins.

17 The use of bioabsorbable materials in medical applications such as the above have the
18 advantages of reducing tissue or cellular irritation and induction of inflammatory response from
19 prominent retained hardware; eliminating or decreasing the necessity of hardware removal; and in
20 the case of orthopedic implants, permitting a gradual stress transfer to the healing bone and thus
21 allowing more complete remodeling of the bone.

22 Bioabsorbable materials for medical applications are well known; for example, United
23 States Patent No. 5,423,859 to Koyfman *et al.*, lists exemplary bioabsorbable or biodegradable
24 resins from which bioabsorbable materials for medical devices may be made. In general,
25 bioabsorbable materials extend to synthetic bioabsorbable, naturally derived polymers, or
26 combinations thereof, with examples as below:

- 27 1) Synthetic Bioabsorbable Polymers: for example polyesters/polylactones such as polymers
28 of polyglycolic acid, glycolide, lactic acid, lactide, dioxanone, trimethylene carbonate etc.,
29 polyanhydrides, polyesteramides, polyorthoesters, polyphosphazenes, and copolymers of
30 these and related polymers or monomers; and

2) Naturally Derived Polymers:

a) Proteins: albumin, fibrin, collagen, elastin;

b) Polysaccharides: chitosan, alginates, hyaluronic acid; and

3) Biosynthetic Polyesters: 3-hydroxybutyrate polymers.

Like other biomaterials, bioabsorbable materials are also subjected to bacterial contamination and can be a source of infections which are difficult to control. Those infections quite often lead to the failure of the devices, requiring their removal and costly antimicrobial treatments.

Prior art efforts to render bioabsorbable materials more infection resistant generally have focused on impregnating the materials with antibiotics or salts such as silver salts. However, such efforts usually provide only limited, and instantaneous antimicrobial activity, which is limited by the availability or solubility of the antimicrobial agent over time. It is desirable to have an antimicrobial effect which is sustained over time, such that the antimicrobial effect can be prolonged for the time that the bioabsorbable material is in place. This can range from hours or days, to weeks or even years.

There are suggestions in the prior art to provide metal coatings, such as silver coatings, on medical devices; for example, International Publication No. WO 92/13491 to Vidal and Redmond; Japanese Patent Application Disclosure No. 21912/85 to Mitsubishi Rayon K.K., Tokyo; and United States Patent No. 4,167,045 to Sawyer. None of these references include teachings specific to the use of metal coatings on bioabsorbable materials. In such applications, it is important that the metal coatings do not shed or leave behind large metal particulates in the body, which will induce unwanted immune responses and/or toxic effects.

There is a need for antimicrobial coatings for bioabsorbable materials, which can create an effective and sustainable antimicrobial effect, which do not interfere with the bioabsorption of the bioabsorbable material, and which do not shed or leave behind large metal particulates in the body as the bioabsorbable material disappears.

SUMMARY OF THE INVENTION

This invention provides bioabsorbable materials comprising a bioabsorbable substrate associated with one or more antimicrobial metals being in a crystalline form characterized by sufficient atomic disorder, such that the bioabsorbable material in contact with an alcohol or water based electrolyte, releases atoms, ion, molecules, or clusters of at least one antimicrobial metal at a

1 concentration sufficient to provide an antimicrobial effect. The one or more antimicrobial metals do
2 not interfere with the bioabsorption of the bioabsorbable material, and do not leave behind
3 particulates larger than $2\text{ }\mu\text{m}$, as measured 24 hours after the bioabsorbable material has
4 disappeared. Most preferably, the particulate sizing from the coating or powder is sub-micron, that
5 is less than about $1\text{ }\mu\text{m}$, as measured 24 hours after the bioabsorbable material has disappeared.
6 Particulates are thus sized to avoid deleterious immune responses or toxic effects. Such
7 antimicrobial metals are in the form of a continuous or discontinuous coating, a powder, or a coating
8 on a bioabsorbable powder.

9 The antimicrobial coating is thin, preferably less than 900 nm or more preferably less than
10 500 nm, and very fine grained, with a grain size (crystallite size) of preferably less than 100 nm,
11 more preferably less than 40 nm, and most preferably less than 20 nm. The antimicrobial coating is
12 formed of an antimicrobial metal, which is overall crystalline, but which is created with atomic
13 disorder, and preferably also having either or both of a) a high oxygen content, as evidenced by a
14 rest potential greater than about 225 mV, more preferably greater than about 250 mV, in 0.15 M
15 Na_2CO_3 against a SCE (standard calomel electrode), or b) discontinuity in the coating.

16 The antimicrobial metal associated with the bioabsorbable substrate may also be in the form
17 of a powder, having a particle size of less than $100\text{ }\mu\text{m}$ or preferably less than $40\text{ }\mu\text{m}$, and with a
18 grain size (crystallite size) of preferably less than 100 nm, more preferably less than 40 nm, and
19 most preferably less than 20 nm. Such powders may be prepared as a coating preferably of the
20 above thickness, onto powdered biocompatible and bioabsorbable substrates; as a nanocrystalline
21 coating and converted into a powder; or as a powder of the antimicrobial metal which is cold
22 worked to impart atomic disorder.

23 A method of preparing the above antimicrobial bioabsorbable materials is also provided,
24 with the bioabsorbable substrate being formed from a bioabsorbable polymer, or being a medical
25 device or part of a medical device. The coating or powder of the one of more antimicrobial metals
26 is formed by either physical vapour deposition under specified conditions and/or by forming the
27 antimicrobial material as a composite material; or by cold working the antimicrobial material
28 containing the antimicrobial metal at conditions which retain the atomic disorder, as in the case
29 where the antimicrobial metal is in the form of a powder. Sufficient oxygen is incorporated in the
30 coating or powder such that particulates of the antimicrobial metals during dissociation are sized at

1 preferably less than 2 μm , or preferably less than 1 μm , to avoid deleterious immune responses or
2 toxic effects.

3 As used herein, the terms and phrases set out below have the meanings which follow.

4 "Alcohol or water-based electrolyte" is meant to include any alcohol or water-based
5 electrolyte that the anti-microbial coatings of the present invention might contact in order to activate
6 (i.e. cause the release of species of the anti-microbial metal) into same. The term is meant to
7 include alcohols, saline, water, gels, fluids, solvents, and tissues containing water, including body
8 fluids (for example blood, urine or saliva), and body tissue (for example skin, muscle or bone).

9 "Antimicrobial effect" means that atoms, ions, molecules or clusters of the anti-microbial
10 metal (hereinafter "species" of the anti-microbial metal) are released into the alcohol or electrolyte
11 which the material contacts in concentrations sufficient to inhibit bacterial (or other microbial)
12 growth in the vicinity of the material. The most common method of measuring anti-microbial effect
13 is by measuring the zone of inhibition (ZOI) created when the material is placed on a bacterial lawn.
14 A relatively small or no ZOI (ex. less than 1 mm) indicates a non useful anti-microbial effect, while a
15 larger ZOI (ex. greater than 5 mm) indicates a highly useful anti-microbial effect. One procedure
16 for a ZOI test is set out in the Examples which follow.

17 "Antimicrobial metals" are metals whose ions have an anti-microbial effect and which are
18 biocompatible. Preferred anti-microbial metals include Ag, Au, Pt, Pd, Ir (i.e., the noble metals),
19 Sn, Cu, Sb, Bi and Zn, with Ag being most preferred.

20 "Atomic disorder" includes high concentrations of: point defects in a crystal lattice,
21 vacancies, line defects such as dislocations, interstitial atoms, amorphous regions, grain and sub grain
22 boundaries and the like relative to its normal ordered crystalline state. Atomic disorder leads to
23 irregularities in surface topography and inhomogeneities in the structure on a nanometer scale.

24 "Bioabsorbable materials" are those useful in medical devices or parts of medical devices,
25 that is which are biocompatible, and which are capable of bioabsorption in a period of time ranging
26 from hours to years, depending on the particular application.

27 "Bioabsorption" means the disappearance of materials from their initial application site in the
28 body (human or mammalian) with or without degradation of the dispersed polymer molecules.

29 "Biocompatible" means generating no significant undesirable host response for the intended
30 utility.

1 “Cold working” as used herein indicates that the material has been mechanically worked
2 such as by milling, grinding, hammering, mortar and pestle or compressing, at temperatures lower
3 than the recrystallization temperature of the material. This ensures that atomic disorder imparted
4 through working is retained in the material.

5 “Diffusion”, when used to describe conditions which limit diffusion in processes to create
6 and retain atomic disorder, i.e. which freeze-in atomic disorder, means diffusion of atoms and/or
7 molecules on the surface or in the matrix of the material being formed.

8 “Dissociation” means the breakdown of the antimicrobial metal in the form of a coating or
9 powder associated with the bioabsorbable substrate, when the bioabsorbable material is in contact
10 with an alcohol or water based electrolyte.

11 “Grain size”, or “crystallite size” means the size of the largest dimension of the crystals in the
12 anti-microbial metal coating or powder.

13 “Metal” or “metals” includes one or more metals whether in the form of substantially pure
14 metals, alloys or compounds such as oxides, nitrides, borides, sulphides, halides or hydrides.

15 “Nanocrystalline” is used herein to denote single-phase or multi-phase polycrystals, the
16 grain size of which is less than about 100, more preferably < 50 and most preferably < 25
17 nanometers in at least one dimension. The term, as applied to the crystallite or grain size in the
18 crystal lattice of coatings, powders or flakes of the anti-microbial metals, is not meant to restrict the
19 particle size of the materials when used in a powder form.

20 “Normal ordered crystalline state” means the crystallinity normally found in bulk metal
21 materials, alloys or compounds formed as cast, wrought or plated metal products. Such materials
22 contain only low concentrations of such atomic defects as vacancies, grain boundaries and
23 dislocations.

24 “Particulate size” means the size of the largest dimension of the particulates which are shed
25 or left behind in the body from the antimicrobial coatings on the bioabsorbable materials.

26 “Powder” is used herein to include particulate sizes of the nanocrystalline anti-microbial
27 metals ranging from nanocrystalline powders to flakes.

28 “Sustained release” or “sustainable basis” are used to define release of atoms, molecules,
29 ions or clusters of an anti-microbial metal that continues over time measured in hours or days, and
30 thus distinguishes release of such metal species from the bulk metal, which release such species at a

1 rate and concentration which is too low to achieve an anti-microbial effect, and from highly soluble
2 salts of anti-microbial metals such as silver nitrate, which releases silver ions virtually instantly, but
3 not continuously, in contact with an alcohol or electrolyte.

4 DESCRIPTION OF THE PREFERRED EMBODIMENTS

5 A. Bioabsorbable Materials

6 Bioabsorbable materials for medical applications are well known, and include
7 bioabsorbable polymers made from a variety of bioabsorbable resins; for example, United States
8 Patent No. 5,423,859 to Koyfman *et al.*, lists exemplary bioabsorbable or biodegradable resins
9 from which bioabsorbable materials for medical devices may be made. Bioabsorbable materials
10 extend to synthetic bioabsorbable or naturally derived polymers, with typical examples as below:

- 11 1) Synthetic Bioabsorbable Polymers: for example polyesters/polylactones such as polymers
12 of polyglycolic acid, glycolide, lactic acid, lactide, dioxanone, trimethylene carbonate etc.,
13 polyanhydrides, polyesteramides, polyorthoesters, polyphosphazenes, and copolymers of
14 these and related polymers or monomers.
- 15 2) Naturally Derived Polymers:
 - 16 a) Proteins: albumin, fibrin, collagen, elastin;
 - 17 b) Polysaccharides: chitosan, alginates, hyaluronic acid; and
- 18 3) Biosynthetic Polyesters: 3-hydroxybutyrate polymers.

19 The bioabsorbable material, depending on the application, may be used in a powder, sheet
20 or fibre form.

21 Many medical applications exist for bioabsorbable materials coated with the antimicrobial
22 coatings of this invention, including, without limitation:

- 23 1) Wound closures: including for example sutures, staples, and adhesives;
- 24 2) Tissue Repair: including for example meshes for hernia repair;
- 25 3) Prosthetic Devices: including for example internal bone fixation, physical barrier for guided
26 bone regeneration;
- 27 4) Tissue Engineering: including for example blood vessels, skin, bone, cartilage, and liver;
- 28 5) Controlled Drug Delivery Systems: including for example microcapsules and ion-exchange
29 resins; and
- 30 6) Wound Coverings or Fillers: including for example alginate dressings and chitosan powders.

1 **B. Antimicrobial Coating for Bioabsorbable Materials**

2 The bioabsorbable material includes an antimicrobial coating formed from an antimicrobial
3 metal, which is formed by the procedure set out below. The coating can be applied as one or more
4 of the layers, but is most preferably applied as a discontinuous coating of a single thin layer which is
5 less than 900 nm in thickness, more preferably less than 500 nm, and which has a grain size (i.e.
6 crystallite size in the coating itself) less than 100 nm, more preferably less than 40 nm, and most
7 preferably less than 20 nm.

8 The coating is most preferably formed with atomic disorder in accordance with the
9 procedures set out above and as described in International Publication Nos. WO 98/41095, WO
10 95/13704, and WO 93/23092, all to Burrell *et al.* In addition, the coating is preferably formed
11 with a high oxygen content, as determined by a positive rest potential greater than 225 mV, more
12 preferably greater than about 250 mV, in 0.15 M Na₂CO₃ against a SCE, when measured in
13 accordance with the procedure set out in Example 5. The high oxygen content is achieved by
14 including oxygen in the working gas atmosphere during the physical vapour deposition technique.
15 Preferably the ratio of inert working gas (preferably argon) to oxygen is about 96:4 or less.

16 The antimicrobial coating can be rendered discontinuous by many techniques, for instance
17 by coating fibers or powders from only one side, with or without rotation or vibration, by making
18 the coatings so thin as to be discontinuous, by coating on porous fibrous materials so as to achieve
19 discontinuity, by masking either the substrate or the cathode, or to etch a continuous coating.

20 The above features of the antimicrobial coatings of this invention have been found to ensure
21 that the particulate size left behind by the antimicrobial coatings as the bioabsorbable material
22 disappears, are less than about 2 μ m in size, and more preferably are less than 1 μ m in size.

23 The antimicrobial coating is formed in a crystalline form from antimicrobial metals with
24 atomic disorder so as to produce an antimicrobial effect. The production of atomic disorder
25 through physical vapour deposition techniques is described in the above mentioned PCT
26 applications to Burrell *et al.* and as outlined below.

27 The antimicrobial metal is deposited as a thin metallic film on one or more surfaces of the
28 bioabsorbable material by vapour deposition techniques. Physical vapour techniques, which are
29 well known in the art, all deposit the metal from the vapour, generally atom by atom, onto a

1 substrate surface. The techniques include vacuum or arc evaporation, sputtering, magnetron
2 sputtering and ion plating. The deposition is conducted in a manner to create atomic disorder in the
3 coating as defined above. Various conditions responsible for producing atomic disorder are useful.
4 These conditions are generally those which one has been taught to avoid in thin film deposition
5 techniques, since the object of most thin film depositions is to create a defect free, smooth and
6 dense film (see for example J.A. Thornton, J. Vac. Sci. Technol., Vol 11, (4); 666-670; and
7 "Coating Deposition by Sputtering" in *Deposition Technologies For Films and Coatings*, Noyes
8 Publications, N.J. 170-237, (1982)). The preferred conditions which are used to create atomic
9 disorder during the deposition process include:

10 - a low substrate temperature, that is maintaining the surface to be coated at a temperature
11 such that the ratio of the substrate temperature to the melting point of the metal (in degrees Kelvin)
12 is less than about 0.5, more preferably less than about 0.35 and most preferably less than about 0.3;
13 and optionally one or both of:

14 - a higher than normal working (or ambient) gas pressure, i.e. for vacuum evaporation: e-
15 beam or arc evaporation, greater than 0.01 mT, gas scattering evaporation (pressure plating) or
16 reactive arc evaporation, greater than 20 mT; for sputtering: greater than 75 mT; for magnetron
17 sputtering: greater than about 10 mT; and for ion plating: greater than about 200 mT; and

18 - maintaining the angle of incidence of the coating flux on the surface to be coated at less
19 than about 75°, and preferably less than about 30°.

20 The metals used in the coating are those known to release ions etc. having an antimicrobial
21 effect, as set out above. For bioabsorbable materials, the metal must also be biocompatible.
22 Preferred metals include the noble metals Ag, Au, Pt, Pd, and Ir as well as Sn, Cu, Sb, Bi, and Zn
23 or alloys or compounds of these metals or other metals. Most preferred is Ag or Au, or alloys or
24 compounds of one or more of these metals. Particularly preferred is Ag.

25 For economic reasons, the thin metal film has a thickness no greater than that needed to
26 provide release of metal ions on a sustainable basis over a suitable period of time. Within the
27 preferred ranges of thicknesses set out above, the thickness will vary with the particular metal in the
28 coating (which varies the solubility and abrasion resistance), and with the degree of atomic disorder
29 in (and thus the solubility of) the coating. The thickness will be thin enough that the coating does not
30 interfere with the dimensional tolerances or flexibility of the device for its intended utility.

1 The antimicrobial effect of the material so produced is achieved when the coating is brought
2 into contact with an alcohol or a water-based electrolyte, thus releasing metal ions, atoms,
3 molecules or clusters. The concentration of the metal species which is needed to produce an
4 antimicrobial effect will vary from metal to metal. Generally, an antimicrobial effect is achieved with
5 silver coatings in body fluids such as plasma, serum or urine at concentrations less than about 0.5 -
6 10 µg/ml of silver species. Evidence of the antimicrobial effect of the material may be demonstrated
7 by biological testing. Localized antimicrobial effect is demonstrated by zone of inhibition testing (see
8 Example 1), whereas sustained release of the antimicrobial metal is illustrated by log reduction (see
9 Examples 2 and 4).

10 The ability to achieve release of metal atoms, ions, molecules or clusters on a sustainable
11 basis from a coating is dictated by a number of factors, including coating characteristics such as
12 composition, structure, solubility and thickness, and the nature of the environment in which the
13 device is used. As the level of atomic disorder is increased, the amount of metal species released
14 per unit time increases. For instance, a silver metal film deposited by magnetron sputtering at T/T_m
15 < 0.5 and a working gas pressure of about 7 mTorr releases approximately 1/3 of the silver ions
16 that a film deposited under similar conditions, but at 30 mTorr, will release over 10 days. Films that
17 are created with an intermediate structure (ex. lower pressure, lower angle of incidence etc.) have
18 Ag release values intermediate to these values as determined by bioassays. This then provides a
19 method for producing controlled release metallic coatings. Slow release coatings are prepared such
20 that the degree of disorder is low while fast release coatings are prepared such that the degree of
21 disorder is high.

22 The time required for total dissolution will be a function of the film thickness, the
23 composition of the film and the nature of the environment to which the film is exposed. The
24 relationship in respect of thickness is approximately linear, i.e., a two-fold increase in film thickness
25 will result in about a two-fold increase in longevity.

26 It is also possible to control the metal release from a coating by forming a thin film coating
27 with a modulated structure. For instance, a coating deposited by magnetron sputtering such that the
28 working gas pressure was low (ex. 15 mTorr) for 50% of the deposition time and high (ex. 30
29 mTorr) for the remaining time, has a rapid initial release of metal ions, followed by a longer period
30 of slow release. This type of coating is extremely effective on devices such as urinary catheters for

1 which an initial rapid release is required to achieve immediate antimicrobial concentrations followed
2 by a lower release rate to sustain the concentration of metal ions over a period of weeks.

3 The substrate temperature used during vapour deposition should not be so low that
4 annealing or recrystallization of the coating takes place as the coating warms to ambient
5 temperatures or the temperatures at which it is to be used (ex. body temperature). This allowable
6 ΔT , that the temperature differential between the substrate temperature during deposition and the
7 ultimate temperature of use, will vary from metal to metal. For the most preferred metals of Ag and
8 Au, preferred substrate temperatures of -20°C to 200°C , more preferably -10°C to 100°C are
9 used.

10 Atomic disorder may also be achieved by preparing composite metal materials, that is
11 materials which contain one or more antimicrobial metals in a metal matrix which includes atoms or
12 molecules different from the antimicrobial metals, such that the inclusion of the different materials
13 creates atomic disorder in the crystalline lattice.

14 The preferred technique for preparing a composite material is to co- or sequentially deposit
15 the antimicrobial metal(s) with one or more other inert, biocompatible metals selected from Ta, Ti,
16 Nb, Zn, V, Hf, Mo, Si, Al and alloys of these metals or other metal elements, typically other
17 transition metals. Such inert metals have a different atomic radii from that of the antimicrobial
18 metals, which results in atomic disorder during deposition. Alloys of this kind can also serve to
19 reduce atomic diffusion and thus stabilize the disordered structure. Thin film deposition equipment
20 with multiple targets for the placement of each of the antimicrobial and inert metals is preferably
21 utilized. When layers are sequentially deposited the layer(s) of the inert metal(s) should be
22 discontinuous, for example as islands within the antimicrobial metal matrix. The final ratio of the
23 antimicrobial metal(s) to inert metal(s) should be greater than about 0.2. The most preferable inert
24 metals are Ti, Ta, Zn and Nb. It is also possible to form the antimicrobial coating from oxides,
25 carbides, nitrides, sulphides, borides, halides or hydrides of one or more of the antimicrobial metals
26 and/or one or more of the inert metals to achieve the desired atomic disorder.

27 Another composite material may be formed by reactively co- or sequentially depositing, by
28 physical vapour techniques, a reacted material into the thin film of the antimicrobial metal(s). The
29 reacted material is an oxide, nitride, carbide, boride, sulphide, hydride or halide of the antimicrobial
30 and/or inert metal, formed in situ by injecting the appropriate reactants, or gases containing same,

1 (ex. air, oxygen, water, nitrogen, hydrogen, boron, sulphur, halogens) into the deposition chamber.
2 Atoms or molecules of these gases may also become absorbed or trapped in the metal film to
3 create atomic disorder. The reactant may be continuously supplied during deposition for
4 codeposition or it may be pulsed to provide for sequential deposition. The final ratio of
5 antimicrobial metal(s) to reaction product should be greater than about 0.2. Air, oxygen, nitrogen
6 and hydrogen are particularly preferred reactants.

7 The above deposition techniques to prepare composite coatings may be used with or
8 without the conditions of lower substrate temperatures, high working gas pressures and low angles
9 of incidence set out above. One or more of these conditions are preferred to retain and enhance
10 the amount of atomic disorder created in the coating.

11 **C. Antimicrobial Powder for Bioabsorbable Materials**

12 Antimicrobial powders for bioabsorbable materials are preferably nanocrystalline powders
13 formed with atomic disorder. The powders either as pure metals, metal alloys or compounds such
14 as metal oxides or metal salts, can be formed by vapour deposition, mechanical working, or
15 compressing to impart atomic disorder, as set out below. Mechanically imparted disorder is
16 conducted under conditions of low temperature (i.e. temperatures less than the temperature of
17 recrystallization of the material) to ensure that annealing or recrystallization does not take place.

18 Nanocrystalline powders may comprise powders of the antimicrobial metal itself, or
19 bioabsorbable powders which are coated with the antimicrobial metal, as demonstrated in Example
20 4 in which a chitosan powder is coated with silver.

21 Nanocrystalline powders of the antimicrobial metals may be prepared by several
22 procedures as set out above, and as described in International Publication Nos. WO 93/23092 and
23 WO 95/13704, both to Burrell *et al.*; or as otherwise known in the art. In general, nanocrystalline
24 powders may be prepared as a nanocrystalline coating (formed with atomic disorder in accordance
25 with procedures previously described) preferably of the above thickness, onto powdered
26 biocompatible and bioabsorbable substrates such as chitin; or may be prepared as a nanocrystalline
27 coating onto a substrate such as a cold finger or a silicon wafer, with the coating then scraped off to
28 form a nanocrystalline powder.

29 Alternatively, fine grained or nanocrystalline powders of the anti-microbial metals may be
30 cold worked to impart atomic disorder, whereby the material has been mechanically worked such

1 as by milling, grinding, hammering, mortar and pestle or compressing, at temperatures lower than
2 the recrystallization temperature of the material to ensure that atomic disorder is retained in the
3 material (International Publication Nos. WO 93/23092 and WO 95/13704, both to Burrell *et al.*).
4 Nanocrystalline powders may be sterilized with gamma radiation as described below to maintain
5 atomic disorder, hence the antimicrobial effect.

6 The prepared nanocrystalline powders may then be incorporated into or onto the
7 bioabsorbable substrate by any methods known in the art. For example, the nanocrystalline
8 powders may be layered onto the bioabsorbable substrate as a coating; mechanically mixed within
9 the fibers of the bioabsorbable substrate; or impregnated into the bioabsorbable substrate by
10 physical blowing. The quantity of nanocrystalline powder impregnating a bioabsorbable substrate
11 could be adjusted accordingly to achieve a desired dose range. Alternatively, the nanocrystalline
12 powder may be incorporated into a polymeric, ceramic, metallic matrix, or other matrices to be
13 used as a material for the manufacture of bioabsorbable substrates, medical devices or parts of
14 medical devices, or coatings therefor.

15 The antimicrobial effect of the nanocrystalline powders is achieved when the substrate,
16 coated or impregnated with the nanocrystalline powder, is brought into contact with an alcohol or a
17 water-based electrolyte, thus releasing the antimicrobial metal ions, atoms, molecules or clusters.

18 **D. Sterilization**

19 Bioabsorbable materials once coated with the antimicrobial coating or powder of an
20 antimicrobial metal formed with atomic disorder are preferably sterilized without applying excessive
21 thermal energy, which can anneal out the atomic disorder, thereby reducing or eliminating a useful
22 antimicrobial effect. Gamma radiation is preferred for sterilizing such dressings, as discussed in
23 International Publication No. WO 95/13704 to Burrell *et al.*

24 The sterilized materials should be sealed in packaging which excludes light penetration to
25 avoid additional oxidation of the antimicrobial coating. Polyester peelable pouches are preferred.
26 The shelf life of bioabsorbable, antimicrobial materials thus sealed should be over one year.

27 **E. Use of Bioabsorbable Materials With Antimicrobial Coating or Powder**

28 The antimicrobial coatings and powders of this invention are activated by contacting an
29 alcohol or water-based electrolyte. If the bioabsorbable material is to be used in an application
30 which does not provide exposure to an electrolyte, the material can be moistened with drops of

1 sterile water or 70% ethanol, in order to activate the coating for release of antimicrobial metal
2 species. In a dressing form, the bioabsorbable material can be secured in place with an occlusive or
3 semi-occlusive layer, such as an adhesive film, which will keep the dressing in a moist environment.

4 **F. Examples**

5 **Example 1 - Silver-coated Bioabsorbable Sutures**

6 **1.1 Bioabsorbable Material**

7 Nanocrystalline silver coating was prepared on a bioabsorbable suture. The bioabsorbable material
8 coated was DEXON™ II BI-COLOR (Braided polyglycolic acid with polycaprolate coating)
9 manufactured by Sherwood Medical Corp. (St. Louis, MO, USA).

10 **1.2 Sputtering Conditions**

11 The coating layer on only one side of the bioabsorbable suture was formed by magnetron sputtering
12 under the following conditions:

13 Target:	99.99% Ag
14 Target Size:	20.3 cm diameter
15 Working Gas:	96/4 wt% Ar/O ₂
16 Working Gas Pressure:	40 mTorr
17 Power:	0.11 kW
18 Substrate Temperature:	20°C
19 Base Pressure:	4.0 X 10 ⁻⁶ Torr
20 Anode/Cathode Distance:	100 mm
21 Sputtering Time/Film Thickness:	16 min/ 500 nm
22 Voltage:	360 V

23 With these sputtering conditions applied to the suture material on only one side, a discontinuous
24 coating which only covers two thirds of the suture surface was achieved. This coating method gave
25 an open circuit potential greater than 225 mV (in Na₂CO₃, against SCE, as in Example 5) and a
26 crystallite size less than 20 nm as confirmed by x-ray diffraction (XRD) test.

27 **1.3 Zone of Inhibition Test:**

28 To establish that silver species were released from the coated bioabsorbable suture and to
29 demonstrate antimicrobial effect, a zone of inhibition test was conducted. Mueller Hinton agar was
30 dispensed into Petri dishes. The agar plates were allowed to dry the surfaces prior to being

inoculated with lawns of *Pseudomonas aeruginosa* ATCC 27317 and *Staphylococcus aureus* ATCC 25923. Immediately after inoculation, the coated suture segments (one inch long) were placed on the center of the plate. The Petri dishes were incubated at 37°C for 24 hours, and the zone of inhibition (ZOI) was measured thereafter. The results showed that the average ZOIs (triplicate samples) were 9.0 mm and 7.6 mm against *Pseudomonas aeruginosa* and *Staphylococcus aureus* respectively. These inhibition zones were remarkable considering the very small diameter (0.38 mm) of the suture.

1.4 Tensile Strength Test

To demonstrate that the silver coating did not inhibit the bioabsorption of the suture, the following tensile strength test was conducted. The suture was cut into segments of 10 inch lengths, and coated with silver using the sputtering conditions mentioned above. The coated and uncoated sutures were placed in beakers containing 50% fetal bovine serum (Gibco/BRL, Life Technologies Corp., Ontario, Canada) in phosphate buffered saline (PBS, pH 7.2). The beakers were incubated at 37°C. Samples were taken out for tensile strength test using Instron Series IX Automated Material Testing System 1.04 (sample rate: 10.00 pts/sec, crosshead speed: 0.500 in/min, humidity: 50%, temperature: 73°F) at days 1, 2 and 4. The percentage of tensile remaining (% = breaking tensile of treated suture/ breaking tensile of untreated suture X 100%) was calculated. The results are shown in the Table 1.

Table 1. Tensile remaining (%) of PBS-Calf serum treated sutures

Sample	Day 1	Day 2	Day 4
Uncoated suture	98.7	96.4	91.8
Silver-coated suture	96.8	93.5	88.2

It will be noted from Table 1, that the silver coatings did not impede the bioabsorption of the suture material, in that the tensile remaining was similar for both uncoated and silver-coated suture.

Example 2 - Silver Coated Bioabsorbable Alginate Wound Dressing

2.1 Bioabsorbable Material

Kaltostat™ calcium-sodium alginate dressing (ConvaTec, Princeton, NJ, USA) was coated with nanocrystalline silver.

2.2 Sputtering Conditions

The coating layer on the bioabsorbable alginate wound dressing was formed by magnetron

1 sputtering under the following conditions:

2 Target:	99.99% Ag
3 Target Size:	20.3 cm diameter
4 Working Gas:	96/4 wt% Ar/O ₂
5 Working Gas Pressure:	40 mTorr
6 Power:	0.10 kW
7 Substrate Temperature:	20°C
8 Base Pressure:	4.0 X 10 ⁻⁶ Torr
9 Anode/Cathode Distance:	100mm
10 Sputtering Time/Film Thickness:	30 min/ 800 nm
11 Voltage:	360 V

12 Because of the discontinuity of the fibers at the surface of the dressing, this coating represented a
13 discontinuous coating.

14 2.3 Bacterial Killing Capacity Test

15 To demonstrate the bactericidal effect of the coated alginate dressing, a bacterial killing capacity test
16 was conducted. The coated alginate dressing was cut into one square inch pieces. *Pseudomonas*
17 *aeruginosa* ATCC 27317 colonies from an overnight culture were inoculated in 5 ml of Tryptic
18 Soy Broth (TSB) and incubated at 37°C until the suspension reached 0.5 McFarland turbidity. 0.5
19 ml of the bacterial suspension were inoculated onto each piece of the dressings and incubated at
20 37°C for two hours. The survival bacteria in the dressing were recovered by vortexing the dressing
21 in 4.5 ml of STS (0.85% sodium chloride, 1% TweenTM 20 and 0.4% sodium thioglycollate)
22 solution. The bacteria in the solution were enumerated by plate counting and the log reduction was
23 calculated. The result showed that the tested silver-coated alginate dressing induced 6.2 log
24 reduction in the two hour test period, thus demonstrating an excellent bacterial killing capacity of the
25 silver-coated alginate dressing.

26 2.4 Evidence of Bioabsorption

27 Silver-coated Kaltostat dressing and uncoated controls (three pieces of each in one square inch)
28 were weighed before testing. Then the dressings were placed in Petri dishes each containing 30 ml
29 of fetal bovine serum (Gibco/BRL, Life Technologies Corp., Ontario, Canada) and incubated at
30 37°C for three days. The dressings were dried in an oven at 60°C overnight and weighed again.

1 Although degradation could be seen in the dishes, the post-weight was higher than pre-weight
2 because the dressing had absorbed a lot of water and formed a gel. For this reason, a relative
3 weight was calculated. The results showed that the relative weights were 1.69 ± 0.18 and $1.74 \pm$
4 0.12 for uncoated control Kaltostat dressing and silver-coated dressing respectively. The
5 difference was not statistically significant.

6 **Example 3 - Double Side Coated Alginate Wound Dressing**

7 **3.1 Bioabsorbable material**

8 Needled calcium alginate fabric was purchased from Acordis Specialty Fibers Corp. (Coventry,
9 UK).

10 **3.2 Sputtering Conditions**

11 The dressing was sputtered on both sides using a four-pass process with two passes for each side.

12 The Westaim Biomedical TMRC unit was used to coat the dressing under the following conditions:

13 Target:	99.99% Ag
14 Target Size:	15.24 cm X 152.4 cm
15 Working Gas:	80/20 wt% Ar/O ₂ - Base coat
16	100/0 wt% Ar/O ₂ - Top coat
17 Working Gas Pressure:	40 mTorr
18 Total current:	81 A for the first and second passes
19	17 A for the third and fourth passes
20 Base Pressure:	5.0×10^{-3} Torr
21 Web Speed:	230 mm/min - Base coat
22	673 mm/min - Top coat
23 Voltage:	430 V - Base coat
24	300 V - Top coat

25 **3.3 Evidence of Biodegradation**

26 Degradation of the double side coated alginate wound dressing in an aqueous solution resulted in an
27 increase of viscosity in that solution. The following test monitored the increase in viscosity as an
28 indicator of biodegradation *in vitro*. The silver coated alginate dressing and uncoated control
29 alginate dressing were cut into 2" x 2" pieces. Four pieces of each dressing (16 square inch in total)
30 were placed in a beaker containing 80 ml of phosphate buffered saline. The beakers were

1 incubated in a shaking incubator at $37 \pm 1^\circ\text{C}$ and 120 ± 5 rpm for 48 ± 2 hours. After vigorously
2 swirling for ten seconds, the solutions were removed for viscosity analysis. The measuring system
3 used was Z1 DIN with a shear rate range from 0 to 2500 l/s.

4 Thirty data points were collected at 60 second intervals. The results are reported and
5 observed as a chart with a shear rate as the x axis and viscosity as the y axis. Since the viscosity of
6 the solution tends to become stabilized after a shear rate of 1000 l/s, three readings of the viscosity
7 at 1400, 1600 and 1800 l/s are averaged to obtain the viscosity of the solution. Such data showed
8 that silver-coated alginate dressing generated an average viscosity of 3.1 cP while control alginate
9 dressing 3.0 cP. These results suggest that both dressings have a very similar degradation rate,
10 which indicates that the silver coating has no significant impact on the degradation of alginate
11 material.

12 **Example 4 - Silver-coated Chitosan Powder**

13 **4.1 Bioabsorbable Material**

14 Chitosan is a partially deacetylated form of chitin, a natural polysaccharide. It can be degraded by
15 lysozyme and absorbed by body. There have been studies shown that it accelerate wound healing
16 in small animals as rats and dogs (Shigemasa Y. *et al.*, Biotechnology and Genetic Engineering
17 Reviews 1995; 13:383-420). The material used for coating was a fine cream-colored chitosan
18 powder purchased from ICN Biomedicals Inc. (Aurora, Ohio, USA).

19 **4.2 Sputtering Conditions**

20 The chitosan powder was coated by magnetron sputtering under the following conditions:

21 Target:	99.99% Ag
22 Target Size:	20.3 cm diameter
23 Working Gas:	80/20 wt% Ar/O ₂
24 Working Gas Pressure:	30 mTorr
25 Power:	0.2 kW
26 Substrate Temperature:	20°C
27 Base Pressure:	6.0×10^{-6} Torr
28 Anode/Cathode Distance:	100 mm
29 Sputtering Time/Film Thickness:	10 min
30 Voltage:	409 V

1 As in Example 1, these coating conditions resulted in a discontinuous coating of silver, estimated at
2 400 - 500 nm thick, being applied from one side only.

3 4.3 Bacterial Killing Capacity Test

4 The test was similar to that used for the Alginate dressing in Example 2 to demonstrate bactericidal
5 ability of the material. The silver-coated chitosan powder samples (0.03 g) were mixed with 0.3 ml
6 of *Pseudomonas aeruginosa* grown in TSB (10^7 cells/ml) and incubated at 37°C for 30 minutes or
7 2 hours. The silver activity was stopped by addition of 2.7 ml of STS solution. The numbers for
8 bacterial survival were determined using standard plate count techniques. The results showed that
9 the silver-coated chitosan powder reduced the number of viable bacteria to undetectable levels both
10 at 30 minutes and 2 hours.

11 Example 5 - X-ray Diffraction and Rest Potential Measurements

12 Samples of the antimicrobial coatings of the present invention were prepared on glass
13 substrates in order to measure the crystallite sizes and the rest potential. The sputtering conditions
14 are set out in Table 2 below. The conditions were similar to those set out in Examples 1 and 2
15 above, but used varying oxygen content in the working gas, as given in Table 2. A comparison
16 coating of pure silver (i.e., sputtered in 100% Ar) was also prepared. The sputtered films were
17 then analyzed by x-ray diffraction to determine the crystallite size, measured for silver along the
18 Ag(111) line, and to estimate for silver oxide by measuring along the Ag₂O(111). The films were
19 also examined electrochemically to determine the rest potential or open circuit potential (OCP).
20 The latter measurement was conducted to confirm a high oxygen content in the films. The rest
21 potential was obtained by two procedures, one being a measurement for 15 minutes in 0.15 M
22 KOH solution and the second being a measurement for 20 minutes in 0.15 M Na₂CO₃ solution,
23 both being against a saturated calomel electrode (SCE). The results are set out in Table 3.

Table 2 - Sputtering Conditions for Samples

Sample number	Sample Ratio Ar:O ₂	Base P mTorr	Gas P mTorr	Current [A]	Voltage [V]	Power [kW]	Dep. Time [min.]	Thick. [nm]
1	100:0	2.3 x 10 ⁻⁶	40±0.5	0.81	345	0.252	10	749
2	96:4	2.7 x 10 ⁻⁶	40±0.4	0.81	400	0.290	10	944
3	94:6	2.5 x 10 ⁻⁶	40±0.3	0.81	410	0.300	10	1120
4	92:8	1.7 x 10 ⁻⁶	40±0.5	0.811	424	0.309	10	1130
5	96:4	3.0 x 10 ⁻⁶	40±0.4	0.320	364	0.107	30	1010

Table 3 - Rest Potentials for Samples under Sputtering Conditions of Table 2

Sample number	Sample Ratio Ar:O ₂	Crystallite Size [nm]	OCP [mV] 0.15 M Na ₂ CO ₃	OCP [mV] 0.15 M KOH
1	100:0	123.9	148	-13
2	96:4	15.4	269	141
3	94:6	10.5	265	138
4	92:8	8.2	259	133
5	96:4	est. as 13-20	>+720	>+650

REFERENCES

- Shigemasa Y. and Minami, S. 1995. Applications of chitin and chitosan for biomaterials. *Biotechnology and Genetic Engineering Reviews* 13: 383-420.
- Thornton, J.A. 1982. Influence of apparatus geometry and deposition conditions on the structure and topography of thick sputtered coatings. *J. Vac. Sci. Technol.* 11(4): 666-670.
- Thornton, J.A. 1982. Coating deposition by sputtering. *Deposition Technologies For Films and Coatings*, Noyes Publications, N.J. pp 170-237.

PATENT DOCUMENTS

- Burrell, R.E., Apte, P.S., McIntosh, C.L., Sant, S.B., Gill, K.S., Morris, L.R., and Precht, R.J. Anti-microbial materials. International Publication No. WO 95/13704, published May 26, 1995.
- Burrell, R.E. and Morris, L.R. Anti-microbial coating for medical devices. International Publication No. WO 93/23092, published November 25, 1993.
- Burrell, R.E. and Precht, R.J. Anti-microbial coatings having indicators and wound dressings. International Publication No. WO 98/41095, published September 24, 1998.
- Koyman, I and Chesterfield, M.P. Jet entangled suture yarn and method for making same. United States Patent No. 5,423,859, issued June 13, 1995.
- Mitsubishi Rayon K.K., Tokyo. Process for the preparation of metal deposition carrying synthetic fibre staples. Japanese Patent Application Disclosure No. 21912/85, published February 4, 1985.
- Sawyer, P.N. Cardiac and vascular prostheses. United States Patent No. 4,167,045, issued September 11, 1979.
- Vidal, C. and Redmond, R.J. Improved surgical hardware with bacteriostatic silver coating, and method of using same. International Publication No. WO 92/13491, published August 20, 1992.

All publications mentioned in this specification are indicative of the level of skill in the art to which this invention pertains. All publications are herein incorporated by reference to the same extent as if each individual publication was specifically and individually indicated to be incorporated by reference.

Although the foregoing invention has been described in some detail by way of illustration

- 1 and example, for purposes of clarity and understanding it will be understood that certain changes
- 2 and modifications may be made without departing from the scope or spirit of the invention as
- 3 defined by the following claims.

1 We claim:

- 2 1. A bioabsorbable material comprising:
3 a bioabsorbable substrate; and
4 one or more antimicrobial metals associated with the bioabsorbable substrate, the one or
5 more antimicrobial metals being in a crystalline form characterized by sufficient atomic
6 disorder, such that the material in contact with an alcohol or water-based electrolyte,
7 releases atoms, ions, molecules, or clusters of at least one antimicrobial metal at a
8 concentration sufficient to provide a localized antimicrobial effect, and wherein the one or
9 more antimicrobial metals are associated with the bioabsorbable substrate such that
10 particulates of the one or more antimicrobial metals formed during dissociation are sized to
11 avoid deleterious immune responses or toxic effects.
- 12 2. The material of claim 1, wherein the one or more antimicrobial metals associated with the
13 bioabsorbable substrate are in the form of a continuous or discontinuous coating or a
14 powder.
- 15 3. The material of claim 1, wherein the one or more antimicrobial metals associated with the
16 bioabsorbable substrate are in the form of a coating on a powder.
- 17 4. The material of claim 2, wherein the one or more antimicrobial metals are formed as
18 discontinuous coatings and/or with sufficient high oxygen content that the particulate of the
19 one or more antimicrobial metals formed during dissociation has a size of less than 2 μm .
- 20 5. The material of claim 4, wherein the particulate has a size of less than 1 μm .
- 21 6. The material of claim 2, wherein the one or more antimicrobial metals are provided in the
22 form of a coating, having a thickness of less than 900 nm.
- 23 7. The material of claim 6, wherein the one or more antimicrobial metals are provided in the
24 form of a coating, having a thickness of less than 500 nm.
- 25 8. The material of claim 2, wherein the one or more antimicrobial metals are provided in the
26 form of a powder, having a particle size of less than 100 μm .
- 27 9. The material of claim 8, wherein the one or more antimicrobial metals are provided in the
28 form of a powder, having a particle size of less than 40 μm .
- 29 10. The material of claim 2, wherein the one or more antimicrobial metals are in the form of a
30 nanocrystalline coating or powder, formed with sufficient atomic disorder to provide

- 1 sustained release of atoms, ions, molecules, or clusters of the one or more antimicrobial
2 metals.
- 3 11. The material of claim 10, wherein the nanocrystalline coating or powder has a crystallite size
4 of less than 100 nm.
- 5 12. The material of claim 10, wherein the nanocrystalline coating or powder has a grain size less
6 than 40 nm.
- 7 13. The material of claim 10, wherein the nanocrystalline coating or powder has a grain size less
8 than 20 nm.
- 9 14. The material of claim 13, wherein the one or more antimicrobial metals are selected from
10 the group consisting of Ag, Au, Pt, Pd, Ir, Sn, Cu, Sb, Bi, Zn, or alloys or compounds
11 thereof.
- 12 15. The material of claim 11, wherein at least one of the one or more antimicrobial metals is Ag
13 or Au, or alloys or compounds thereof.
- 14 16. The material of claim 11, wherein the antimicrobial metal is silver, or an alloy or compound
15 thereof.
- 16 17. The material of claim 14, wherein the coating or powder includes absorbed, trapped, or
17 reacted atoms or molecules of oxygen.
- 18 18. The material of claim 17, wherein sufficient oxygen is incorporated in the coating or powder
19 such that the particulate of the one or more antimicrobial metals during dissociation has a
20 size less than 2 μm .
- 21 19. The material of claim 17, wherein sufficient oxygen is incorporated in the coating or powder
22 such that the particulate of the one or more antimicrobial metals during dissociation has a
23 size less than 1 μm .
- 24 20. The material of claim 18, wherein the one or more antimicrobial metals are silver, or an alloy
25 or compound thereof, and wherein the coating or powder has a ratio of its temperature of
26 recrystallization to its melting temperature, in degrees K ($T_{\text{rec}}/T_{\text{m}}$), less than 0.33.
- 27 21. The material of claim 20, wherein the ratio is less than 0.3.
- 28 22. The material of claim 21, wherein the temperature of recrystallization is less than about
29 140°C.
- 30 23. The material of claim 20, wherein the coating has a positive rest potential, when measured

- 1 against a standard calomel electrode, in 0.15 M Na₂CO₃ or 0.15 M KOH.
- 2 24. The material of claim 23, wherein the positive rest potential is greater than 225 mV in
3 0.15 M Na₂CO₃.
- 4 25. The material of claim 23, wherein the positive rest potential is greater than 250 mV in
5 0.15 M Na₂CO₃.
- 6 26. The material of claim 23, wherein the bioabsorbable substrate is formed from a
7 bioabsorbable polymer selected from:
- 8 (a) polyester or polylactone selected from the group comprising polymers of
9 polyglycolic acid, glycolide, lactic acid, lactide, dioxanone, trimethylene carbonate,
10 polyanhydrides, polyesteramides, polyorthoesters, polyphosphazenes, and
11 copolymers of these and related polymers or monomers;
- 12 (b) protein, selected from the group comprising albumin, fibrin, collagen, or elastin;
- 13 (c) polysaccharide, selected from the group comprising chitosan, alginates, or
14 hyaluronic acid; or
- 15 (d) biosynthetic polymer, comprising 3-hydroxybutyrate polymers.
- 16 27. The material of claim 23, wherein the bioabsorbable substrate is a medical device or a part
17 of a medical device selected from: a wound closure, a suture, a staple, an adhesive; a
18 mesh; a prosthetic device; a controlled drug delivery system; a wound covering; and a filler.
- 19 28. The material of claim 23, wherein the bioabsorbable substrate is an alginate dressing coated
20 with a coating of the one or more antimicrobial metals or impregnated with a powder of the
21 one or more antimicrobial metals.
- 22 29. The material of claim 23, wherein the bioabsorbable substrate is a chitosan powder coated
23 with a coating of the one or more antimicrobial metals.
- 24 30. A method of preparing a bioabsorbable material comprising:
25 providing a bioabsorbable substrate; and
26 contacting the bioabsorbable substrate with one or more antimicrobial metals, such that the
27 one or more antimicrobial metals remain associated with the bioabsorbable substrate, the
28 one of more antimicrobial metals being formed by creating atomic disorder under process
29 conditions which limit diffusion for retaining atomic disorder therein, the atomic disorder
30 being sufficient, such that the material in contact with an alcohol or water-based electrolyte,

- 1 releases atoms, ions, molecules, or clusters of at least one anti-microbial metal at a
2 concentration sufficient to provide a localized antimicrobial effect, and wherein the one or
3 more antimicrobial metals are associated with the bioabsorbable substrate such that
4 particulates of the one or more antimicrobial metals formed during dissociation are sized to
5 avoid deleterious immune responses or toxic effects.
- 6 31. The method of claim 30, wherein the bioabsorbable substrate is contacted with the one or
7 more antimicrobial metals by forming a coating on the bioabsorbable substrate, or by
8 incorporating a powder into or onto the bioabsorbable substrate, and wherein the one or
9 more antimicrobial metals associated with the bioabsorbable substrate are formed as a
10 continuous or discontinuous coating or powder.
- 11 32. The method of claim 30, wherein the one or more antimicrobial metals associated with the
12 bioabsorbable substrate are formed as a coating on a powder.
- 13 33. The method of claim 31, wherein the one or more antimicrobial metals are formed as
14 discontinuous coatings and/or with sufficient high oxygen content that the particulate of the
15 one or more antimicrobial metals formed during dissociation has a size of less than 2 μm .
- 16 34. The method of claim 33, wherein the particulate has a size of less than 1 μm .
- 17 35. The method of claim 31, wherein the one or more antimicrobial metals are formed as a
18 coating, having a thickness of less than 900 nm.
- 19 36. The method of claim 35, wherein the one or more antimicrobial metals are formed as a
20 coating, having a thickness of less than 500 nm.
- 21 37. The method of claim 31, wherein the one or more antimicrobial metals are formed as a
22 powder, having a particle size of less than 100 μm .
- 23 38. The method of claim 37, wherein the one or more antimicrobial metals are formed as a
24 powder, having a particle size of less than 40 μm .
- 25 39. The method of claim 31, wherein the one or more antimicrobial metals are formed as a
26 nanocrystalline coating or powder, formed with sufficient atomic disorder to provide
27 sustained release on atoms, ions, molecules, or clusters of the one or more antimicrobial
28 metals.
- 29 40. The method of claim 39, wherein the one or more antimicrobial metals are formed as a
30 nanocrystalline coating or powder having a crystallite size of less than 100 nm.

- 1 41. The method of claim 40, wherein the one or more antimicrobial metals are formed as a
2 nanocrystalline coating or powder having a grain size less than 40 nm.
- 3 42. The method of claim 41, wherein the one or more antimicrobial metals are formed as a
4 nanocrystalline coating or powder having a grain size less than 20 nm.
- 5 43. The method of claim 40, wherein the one or more antimicrobial metals are selected from the
6 group consisting of Ag, Au, Pt, Pd, Ir, Sn, Cu, Sb, Bi, and Zn, or alloys or compounds
7 thereof.
- 8 44. The method of claim 40, wherein at least one of the one or more antimicrobial metal is Ag
9 or Au, or alloys or compounds thereof.
- 10 45. The method of claim 40, wherein the antimicrobial metal is silver, or an alloy or compound
11 thereof.
- 12 46. The method of claim 43, wherein the coating or powder of the one or more antimicrobial
13 metals is formed by either
- 14 A. physical vapor deposition selected from vacuum evaporation, sputtering, magnetron
15 sputtering, or ion plating, under one or more of the following conditions:
- 16 (a) maintaining the ratio of the temperature of the substrate being coated to the melting
17 point of the one of more antimicrobial metals or metal compounds being deposited,
18 in degrees Kelvin, at less than 0.5;
- 19 (b) maintaining the angle of incidence of the coating flux on the surface to be coated at
20 less than 75°; and
- 21 (c) maintaining ambient or working gas pressures, depending on the technique of
22 vapour deposition, of:
- 23 (i) greater than 0.01 mT, if by e-beam or arc evaporation;
- 24 (ii) greater than 20 mT, if by gas scattering or reactive arc evaporation;
- 25 (iii) greater than 75 mT, if by sputtering;
- 26 (iv) greater than 10 mT, if by magnetron sputtering; or
- 27 (v) greater than 200 mT, if by ion plating;
- 28 or
- 29 B. by forming the antimicrobial material as a composite material containing the one or
30 more antimicrobial metals by co-, sequentially or reactively depositing by vapour

- 1 deposition, an anti-microbial metal in a crystalline matrix with atoms or molecules of
2 a material different from the antimicrobial metal, the atoms or molecules of the
3 different material creating atomic disorder in the matrix;
- 4 or
- 5 C. cold working an antimicrobial material containing the one or more antimicrobial
6 metals at a temperature below the recrystallization temperature for the material to
7 retain the atomic disorder, wherein the antimicrobial metal is in the form of a
8 powder.
- 9 47. The method of claim 46, wherein the antimicrobial metal is formed as a coating under the
10 conditions set forth in step A and/or B, and includes absorbed, trapped, or reacted atoms
11 or molecules of oxygen which is included in the working gas atmosphere during deposition,
12 said coating being either formed directly on the bioabsorbable substrate, or as a coating
13 which is converted to a powder.
- 14 48. The method of claim 47, wherein a ratio of the working gas to oxygen is about 96:4 or less.
- 15 49. The method of claim 47, wherein sufficient oxygen is incorporated in the coating or powder
16 such that the particulate of the one or more antimicrobial metals during dissociation has a
17 size less than 2 μm .
- 18 50. The method of claim 49, wherein the particulate of the one or more antimicrobial metals
19 during dissociation has a size less than 1 μm .
- 20 51. The method of claim 49, wherein the one or more antimicrobial metals is silver, or an alloy
21 or compound thereof, and wherein the coating or powder has a ratio of its temperature of
22 recrystallization to its melting temperature, in degrees K ($T_{\text{rec}}/T_{\text{m}}$), less than 0.33.
- 23 52. The method of claim 51, wherein the ratio is less than 0.3.
- 24 53. The method of claim 52, wherein the temperature of recrystallization is less than about
25 140°C.
- 26 54. The method of claim 51, wherein the coating has a positive rest potential, when measured
27 against a standard calomel electrode, in 0.15 M Na_2CO_3 or 0.15 M KOH.
- 28 55. The method of claim 54, wherein the positive rest potential is greater than 225 mV in
29 0.15 M Na_2CO_3 .
- 30 56. The method of claim 54, wherein the positive rest potential is greater than 250 mV in 0.15

- 1 M Na₂CO₃.
- 2 57. The method of claim 54, wherein the bioabsorbable substrate is formed from a
- 3 bioabsorbable polymer selected from:
- 4 (a) polyester or polylactone selected from the group comprising polymers of
- 5 polyglycolic acid, glycolide, lactic acid, lactide, dioxanone, trimethylene carbonate,
- 6 polyanhydrides, polyesteramides, polyorthoesters, polyphosphazenes, and
- 7 copolymers of these and related polymers or monomers;
- 8 (b) protein, selected from the group comprising albumin, fibrin, collagen, or elastin;
- 9 (c) polysaccharide, selected from the group comprising chitosan, alginates, or
- 10 hyaluronic acid; or
- 11 (d) biosynthetic polymer, comprising 3-hydroxybutyrate polymers.
- 12 58. The method of claim 54, wherein the bioabsorbable substrate is a medical device or a part
- 13 of a medical device selected from: a wound closure, a suture, a staple, an adhesive; a
- 14 mesh; a prosthetic device; a controlled drug delivery system; a wound covering; and a filler.
- 15 59. The method of claim 54, wherein the bioabsorbable substrate is an alginate dressing coated
- 16 with a coating of the one or more antimicrobial metals or impregnated with a powder of the
- 17 one or more antimicrobial metals.
- 18 60. The material of claim 54, wherein the bioabsorbable substrate is a chitosan powder coated
- 19 with a coating of the one or more antimicrobial metals.